

Low Level laser Therapy (LLLT) / Photobiomodulation (PBMT)

Prepared by James D Carroll

THOR Photomedicine Ltd (UK)

www.thorlaser.com

More information available online

- WEB <http://thorlaser.com>
- UNITED NATIONS TALK [here](#)
- NovoTHOR [here](#)
- RESEARCH [here](#)
- VIDEO INTERVIEWS [here](#)
- BLOG [here](#)
- GET TRAINED [here](#)
- LX2 BROCHURE [here](#)
- Sign up to THOR LLLT Newsletter [here](#)

INTRODUCTION TO PHOTOBIO-MODULATION (PBMT)

WHAT IS PBMT?

Photobiomodulation (Formerly known as Low Level Light / Laser Therapy (LLLT)) is the application of light (usually a low power laser or LED) to promote tissue repair, reduce inflammation or induce analgesia. LLLT has been the subject of several systematic reviews for a range of musculoskeletal pathologies with favorable conclusions reported by The Lancet [1], BMJ [2], International Association for the Study of Pain [3] and the World Health Organization [4]. Unlike other many treatments LLLT is not an ablating or heating based therapy, it is more akin to photosynthesis.

The LLLT devices are typically in the red and near infrared spectrum. Sometimes pulsed and sometimes continuous beams are used. For acute and post operative pathologies as little as one session is all that is necessary but for chronic pain and degenerative conditions as many as ten sessions may be necessary. Whilst other wavelengths have similar effects they do not penetrate nearly so well as the red and near infrared range [5].

RESEARCH:

To-date more than 400 randomized double blind placebo controlled clinical trials have been published with some professional guidelines suggesting LLLT is used as part of standard care, including :

World Health Organization (WHO) Task Force on Neck Pain systematic review [4]

The Lancet Systematic review of LLLT for Neck Pain [1]

International Association for the Study of Pain (IASP) fact sheets for Myofascial Pain Syndrome , osteoarthritis and neck pain [3]

British Medical Journal (BMJ) Systematic review and guidelines for treating tennis elbow [2]

American Physical Therapy Association (APTA) Systematic review and clinical practice guidelines for achilles tendinopathy: [8]

British Journal of Sports Medicine (BJSM) Systematic review for frozen shoulder [7]

European Society for Medical Oncology (ESMO) Clinical practice guidelines for oral mucositis [9]

Multinational Association for Supportive Cancer Care (MASCC) Clinical practice guidelines for oral mucositis [10]

APPLICATIONS

- Arthropathies
- Degenerative Disc Disease
- Fractures
- Lymphedema
- Myofascial pain
- Neuropathies
- Rehabilitation
- Soft tissue injuries
- Tendinopathies

MECHANISM OF ACTION

Most of the effects of LLLT can be explained by light absorption in the mitochondria [14-16]. Every cell in the body has lots of mitochondria (hundreds or thousands per cell). Mitochondria to make cellular energy (ATP) from oxygen and pyruvate. In stressed or ischemic tissues, mitochondria make their own nitric oxide (mtNO) [17-19] which competes with oxygen. The mtNO binds to Cytochrome c Oxidase (CcO) (the terminal enzyme in the electron transport chain) and displaces oxygen [20]. This displacement of oxygen has two negative effects; Reduced ATP synthesis

Increased oxidative stress (leading to inflammation via the inflammatory “master switch” NF-kB) [17-19, 21-23]. The effect of LLLT on hypoxic / stressed tissues can be described in several stages:

Primary effect of LLLT: Absorption by cytochrome c oxidase

Cytochrome c oxidase (CcO) absorbs red and near infrared light, the transfer of light energy by this enzyme triggers a series of downstream effects [14, 24-26].

Secondary effect: Modulation of ATP, nitric oxide & reactive oxygen species

Changes in ATP, reactive oxygen species and nitric oxide follow light absorption by CcO. These effects are redox state and dose dependent. In hypoxic or otherwise stressed cells it has been shown many times that following LLLT, nitric oxide is released, ATP is increased and oxidative stress is reduced [27-31].

Tertiary effect: Downstream intracellular responses (gene transcription, and cellular signaling)

The downstream effects of LLLT released nitric oxide, increased ATP and reduced oxidative stress are many. They are context and cell type specific. Either directly or indirectly these biochemical intermediates affect components in the cytosol, cell membrane, and nucleus that control gene transcription and subsequently cell proliferation, migration, necrosis and inflammation [27-31].

Quaternary effect: Extracellular, indirect, distant effects

Tissues that have not absorbed photons can also be affected indirectly via secretions from cells that have absorbed light. Cells in blood and lymph can be activated and they travel significant distances from the session area to have distant (systemic) effects [32]. These can be autocrine, paracrine, and endocrine effects (sometimes known as a “bystander” effects).

Edema / Lymphatic flow

There is good evidence that LLLT also improves lymphatic flow. A systematic review of eight clinical trials of LLLT for post mastectomy lymphoedema concludes that “There is moderate to strong evidence for the effectiveness of LLLT for the management of breast cancer related lymphedema” [33]. A controlled clinical trial on soccer players with second degree ankle sprains, found a significant reduction in edema volume for the laser group compared with placebo laser (both groups also had rest, ice, compression and elevation) [34]. A laboratory trial on Carrageenan-induced edema in the mouse paw found that treating lymph nodes alone was enough to reduce edema in the mouse paw [35]. The mechanism of action is unknown.

Analgesia

Analgesic effects are probably via a different mechanism from the increased ATP / reduced oxidative stress model described above. According to a systematic review of laser analgesia mechanisms by Chow et al [36], higher power density laser light > 300mW/cm², when absorbed by nociceptors, have an inhibitory effect on A δ and C pain fibers. This high power density LLLT session slows conduction velocity, reduces amplitude of compound action potentials and suppresses neurogenic inflammation. Chow’s own laboratory studies show that LLLT blocks anterograde transport of ATP-rich mitochondria in dorsal root ganglion neurons. Varicosities result from this inhibition, this is normally associated with disruption of microtubules. This effect is completely reversible and lasts only 48 hours [37-39]. More work is needed to fully understand the complete mechanism of action.

Myofascial Trigger points

Myofascial trigger points are palpable nodules in taut muscle bands and contraction of muscle fibers that lead to muscle spasms and limited joint movement. They are a component of several pain conditions, including migraine, tension-type headaches, temporomandibular disorder and neck pain. The motor end plate is central to the etiology of trigger points and EMG studies have shown abnormally high electrical activity over trigger points. Electrical activity is reduced after after LLLT and clinical studies have shown

that LLLT has immediate and cumulative effects on reducing pain [40-43], however the mechanism of action is not yet fully understood.

LLLT PARAMETERS

For LLLT to be effective, the irradiation parameters (wavelength, power, power density, pulse parameters) need to be within certain ranges. If the wrong irradiation parameters are used or applied for the wrong amount of time, then session will be ineffective. If the power density is too low and / or the time is too short, then there is no significant effect. Alternatively, if the power density is too high and / or the session time is too long, then the benefit is lost and sometimes unwanted inhibitory effects occur [44-46].

Unfortunately, many researchers fail to accurately measure or even report some of these parameters. This is due, in part, to a poor appreciation of the relevance of these parameters and also because some of these measurements require expensive instrumentation that need to be operated by trained engineers or physicists [47].

Parameters should be considered in two parts: the 'medicine' and the 'dose'.

MEDICINE

Wavelength (nm)

It is the structure of cytochrome c oxidase and its redox state that determines which wavelengths of light will be absorbed [14-16]. The optimum wavelength is not universally agreed, but most common LLLT devices used in dentistry are typically within the 600nm - 1000nm range. There are many absorption peaks for cytochrome c oxidase in that range, they penetrate tissues well, and many clinical trials have been successful with them.

Power (W)

The most common LLLT devices used in dentistry are in the range 50 - 200mW, but power density is just as important (if not more so), especially for large beam areas.

Beam Area (cm²)

Beam area is required for calculating power density, but is difficult to measure and frequently misreported. Diode laser beams are typically not round (more often they are like an ellipse) and the beams are usually brighter in the middle and gradually weaken towards the edge (Gaussian distribution). This has been poorly understood by many researchers, errors are frequently made when reporting beam area. The aperture does not necessarily define the beam size, which should be measured using a beam profiler and reported at the 1/e² point [47, 96].

Power Density (W/cm²)

Power density is the product of Power (W)/beam area (cm²). This parameter is frequently misreported due to difficulty with measuring beam area [47, 68]. Studies that have taken the trouble to measure beam power density carefully and taken measurements at the target depth report successful tissue repair and anti-inflammatory effects in the range of 5 - 55mW/cm² at the target [65-67]. Analgesia typically requires higher power densities; a systematic review of laboratory studies found power densities > 300 mW/cm² are necessary to inhibit nerve conduction in C-fibres and A-delta fibres [36]

Pulse Structure (Peak Power (W), Pulse freq. (Hz), Pulse Width (s), Duty cycle (%))

If the beam is pulsed, then the reported power should be the "Average Power" and calculated as follows: Peak Power (W) × pulse width (s) × pulse frequency (Hz) = Average Power (W). A review of the effect of pulses [64] concludes that "there was some evidence that pulsed light does have effects that are different from those of continuous wave light. However further work is needed to define these effects for different disease conditions and pulse structures." A subsequent study on traumatic brain injury in mice [63] showed

that 10Hz to be more effective than 100Hz or CW in reducing the neurological severity score.

Coherence

Coherent light produces laser speckle, which has been postulated to play a role in the photobiomodulation interaction with cells and sub-cellular organelles. No definitive trials have been published to-date to confirm or refute this claim but it is clear that coherence is not required to have good clinical effects. [5]

DOSE

Having established suitable irradiation parameters, they must be applied for adequate amount of time. If the wrong irradiation parameters are used or applied for the wrong irradiation time, then session will be ineffective [44, 45, 48, 49].

Energy (Joules) or energy density (fluence) (W/cm²) is often referred to as "dose". These are different calculations and, on their own, are both potentially flawed methods of reporting this therapy. Table 2 shows the formulas and discusses the limitations.

Energy (J)

Calculated as: Power (W) x time (s) = Energy (Joules). Using Joules as an expression of dose is potentially unreliable as it assumes an inverse relationship between power and time and ignores power density.

Energy Density (J/cm²)

Calculated as: Power (W) x time (s) / beam area = Energy Density (J/cm²). Using energy density as an expression of dose is also potentially unreliable, as it assumes an inverse relationship between power, time and power density.

Irradiation time (s)

Given the lack of reciprocity described above, the safest way to record and prescribe LLLT is to define the irradiation parameters, then define the irradiation time and not rely on the total energy or fluence applied.

Treatment Interval (hours, days, or weeks)

One session of acute injuries (or immediately post op) has clinically meaningful effects (though follow up session the next day may also be welcomed by the patient). For chronic non healing or chronic pain pathologies, LLLT typically requires two or three sessions a week for several weeks to achieve clinical significance.

TREATMENTS

There are four common clinical targets for LLLT.

- The site of injury to promote healing, remodeling and reduce inflammation [52-56]
- Lymph nodes to help reduce edema and inflammation. [33, 35, 57]
- Nerves to induce analgesia [36, 37, 39, 58]
- Trigger points to reduce tenderness and relax contracted muscle fibers [40-43].

SAFETY

LED products are considered safe and excluded from medical surveillance under Federal Law 21CFR 1040.10.

Precautions

The North American Association for Laser Therapy conference 2010 held a consensus meeting on safety and contraindications: The main recommendations included:

CANCER - Do not treat over the site of any known primary carcinoma or secondary metastasis unless the patient is undergoing chemotherapy; it's use can be considered in terminally ill cancer patients for palliative relief.

PREGNANCY - Do not treat directly over a developing fetus (consequences unknown)

EPILEPTICS - Be aware that low frequency pulsed visible light (<30hz) might trigger a seizure in photosensitive epileptic patients. Make sure they cannot see any pulsing beams.

Post Treatment Reactions

The Lancet review on neck pain [1] reported that "Half the studies obtained data for side-effects, with tiredness reported in the laser-treated group in three studies,42'46" which was significant in one study"

A chronic joint disorder systematic review [41] reported: "In terms of side effects, six of the LLLT trials with optimal dose explicitly stated in their report that no adverse effects were observed. One trial reported an incident of transient adverse effects for one patient in each group.

CONCLUSION

LLLT is a safe effective session for faster healing, better tissue remodeling, reduced inflammation and analgesia in a wide range of oral pathologies. It is drug free and relatively side-effect free and seems to work where pharmaceuticals do not [11-13, 60-62].

GLOSSARY

Beam profiler

An instrument for measuring the beam intensity distribution

Laser speckle

A random fuzzy looking pattern produced by coherent laser light. Technically speaking they are a random intensity pattern produced by the mutual interference of a set of wavefronts

LED

Light Emitting Diode. A narrow spectral width (one colour) semiconductor light source.

Off-Label

Use for a condition other than that for which it has been officially approved by a regulatory authority (e.g. FDA in USA, CE for Europe, Health Canada, TGA in Australia).

Systematic Review

A review in which research about a topic has been systematically identified, appraised and summarized.

Tissue Remodeling

The third phase of tissue repair after inflammation and cell proliferation

ABOUT THE AUTHOR

JAMES CARROLL

- CEO of THOR Photomedicine Ltd www.thorlaser.com.
- A recognized authority on LLLT mechanisms of action, dose, dose rate effects and the measurement and reporting of parameters.
 - [Scopus Author ID: 55437132700](#)
- Collaborates with universities on LLLT protocol design and the reporting of session parameters:
 - Harvard Medical School
 - Harvard School of Public Health
 - Massachusetts General Hospital
 - Brigham and Women's Hospital (Boston)
 - Massachusetts Institute of technology (MIT)
 - University of Birmingham School of Dentistry
 - Leiden University Medical Centre, Amsterdam
- Appointments :
 - Founder and CEO at THOR Photomedicine Ltd.
 - Biomedical Optics Society conference chair (2009 - to-date)
 - Fellow of The Royal Society of Medicine (2009 - to-date)
 - Editorial Board of Photomedicine and Laser Surgery (2008 to-date)
 - World Association for Laser Therapy (WALT) board member (2000 - 2004)
 - North American Association for Laser Therapy board member (2002 - 2006)

REFERENCES:

1. Chow, R.T., et al., *Efficacy of low-level laser therapy in the management of neck pain: a systematic review and meta-analysis of randomised placebo or active-session controlled trials*. *Lancet*, 2009. **374**(9705): p. 1897-908.
2. Bisset, L., B. Coombes, and B. Vicenzino, *Tennis elbow*. Clinical evidence, 2011. **2011**.
3. IASP, *Global Year Against Musculoskeletal Pain*. <http://tinyurl.com/IASPlaser>, 2010.
4. Haldeman, S., et al., *The Bone and Joint Decade 2000-2010 Task Force on Neck Pain and Its Associated Disorders: executive summary*. *J Manipulative Physiol Ther*, 2009. **32**(2 Suppl): p. S7-9.
5. Chung, H., et al., *The Nuts and Bolts of Low-level Laser (Light) Therapy*. *Annals of biomedical engineering*, 2011.
6. Mester, E., b. Szende, and J.G. Tota, *Effect of laser on hair growth of mice*. *Kiserl Orvostud*, 1967. **19**: p. 628-631.
7. Favejee, M.M., B.M. Huisstede, and B.W. Koes, *Frozen shoulder: the effectiveness of conservative and surgical interventions--systematic review*. *Br J Sports Med*. **45**(1): p. 49-56.
8. Carcia, C.R., et al., *Achilles pain, stiffness, and muscle power deficits: achilles tendinitis*. *The Journal of orthopaedic and sports physical therapy*, 2010. **40**(9): p. A1-26.
9. Peterson, D.E., R.J. Bensadoun, and F. Roila, *Management of oral and gastrointestinal mucositis: ESMO Clinical Practice Guidelines*. *Annals of oncology : official journal of the European Society for Medical Oncology / ESMO*, 2011. **22 Suppl 6**: p. vi78-84.
10. Migliorati, C., et al., *Systematic review of laser and other light therapy for the management of oral mucositis in cancer patients*. *Supportive care in cancer : official journal of the Multinational Association of Supportive Care in Cancer*, 2012.
11. Marcos, R.L., et al., *Infrared (810 nm) low-level laser therapy in rat achilles tendinitis: a consistent alternative to drugs*. *Photochemistry and photobiology*, 2011. **87**(6): p. 1447-52.
12. Bjordal, J.M., et al., *Photoradiation in acute pain: a systematic review of possible mechanisms of action and clinical effects in randomized placebo-controlled trials*. *Photomedicine and laser surgery*, 2006. **24**(2): p. 158-68.
13. Xiaoting, L., T. Yin, and C. Yangxi, *Interventions for pain during fixed orthodontic appliance therapy. A systematic review*. *The Angle orthodontist*, 2010. **80**(5): p. 925-32.
14. Karu, T.I., *Mitochondrial signaling in mammalian cells activated by red and near-IR radiation*. *Photochem Photobiol*, 2008. **84**(5): p. 1091-9.
15. Eells, J.T., et al., *Mitochondrial signal transduction in accelerated wound and retinal healing by near-infrared light therapy*. *Mitochondrion*, 2004. **4**(5-6): p. 559-67.
16. Karu, T., *Mitochondrial mechanisms of photobiomodulation in context of new data about multiple roles of ATP*. *Photomedicine and Laser Surgery*, 2010. **28**(2): p. 159-60.
17. Palacios-Callender, M., et al., *Endogenous NO regulates superoxide production at low oxygen concentrations by modifying the redox state of cytochrome c oxidase*. *Proceedings of the National Academy of Sciences of the United States of America*, 2004. **101**(20): p. 7630-5.
18. Cleeter, M.W., et al., *Reversible inhibition of cytochrome c oxidase, the terminal enzyme of the mitochondrial respiratory chain, by nitric oxide. Implications for neurodegenerative diseases*. *FEBS letters*, 1994. **345**(1): p. 50-4.
19. Antunes, F., A. Boveris, and E. Cadenas, *On the mechanism and biology of cytochrome oxidase inhibition by nitric oxide*. *Proc. Natl. Acad. Sci. USA*, 2004. **101**: p. 16774-9.
20. Galkin, A., A. Higgs, and S. Moncada, *Nitric oxide and hypoxia*. *Essays in biochemistry*, 2007. **43**: p. 29-42.
21. Lane, N., *Cell biology: power games*. *Nature*, 2006. **443**(7114): p. 901-3.
22. Bolanos, J.P., et al., *Nitric oxide-mediated inhibition of the mitochondrial respiratory chain in cultured astrocytes*. *Journal of neurochemistry*, 1994. **63**(3): p. 910-6.

23. Chen, S., *Natural products triggering biological targets--a review of the anti-inflammatory phytochemicals targeting the arachidonic acid pathway in allergy asthma and rheumatoid arthritis*. Current drug targets, 2011. **12**(3): p. 288-301.
24. Karu, T.I. and S.F. Kolyakov, *Exact action spectra for cellular responses relevant to phototherapy*. Photomed Laser Surg, 2005. **23**(4): p. 355-61.
25. Yu, W., et al., *Photomodulation of oxidative metabolism and electron chain enzymes in rat liver mitochondria*. Photochemistry and photobiology, 1997. **66**(6): p. 866-71.
26. Dyson, M., *Primary, secondary and tertiary effects of phototherapy*. Proc. SPIE 6140, Mechanisms for Low-Light Therapy, 614005, 2006.
27. Zhang, R., et al., *Near infrared light protects cardiomyocytes from hypoxia and reoxygenation injury by a nitric oxide dependent mechanism*. Journal of molecular and cellular cardiology, 2009. **46**(1): p. 4-14.
28. Lim, W., et al., *Modulation of Lipopolysaccharide-Induced NF-kappaB Signaling Pathway by 635 nm Irradiation via Heat Shock Protein 27 in Human Gingival Fibroblast Cells*. Photochemistry and photobiology, 2012.
29. Sharma, S.K., et al., *Dose response effects of 810 nm laser light on mouse primary cortical neurons*. Lasers in surgery and medicine, 2011. **43**(8): p. 851-9.
30. de Lima, F.M., et al., *Low-Level Laser Therapy Restores the Oxidative Stress Balance in Acute Lung Injury Induced by Gut Ischemia and Reperfusion*. Photochemistry and photobiology, 2012.
31. Servetto, N., et al., *Evaluation of inflammatory biomarkers associated with oxidative stress and histological assessment of low-level laser therapy in experimental myopathy*. Lasers in surgery and medicine, 2010. **42**(6): p. 577-83.
32. Hopkins, J.T., et al., *Low-Level Laser Therapy Facilitates Superficial Wound Healing in Humans: A Triple-Blind, Sham-Controlled Study*. J Athl Train, 2004. **39**(3): p. 223-229.
33. Omar, M.T., A.A. Shaheen, and H. Zafar, *A systematic review of the effect of low-level laser therapy in the management of breast cancer-related lymphedema*. Supportive care in cancer : official journal of the Multinational Association of Supportive Care in Cancer, 2012. **20**(11): p. 2977-84.
34. Stergioulas, A., *Low-level laser session can reduce edema in second degree ankle sprains*. Journal of clinical laser medicine & surgery, 2004. **22**(2): p. 125-8.
35. Meneguzzo, D.T., et al., *Prevention and session of mice paw edema by near-infrared low-level laser therapy on lymph nodes*. Lasers in medical science, 2012.
36. Chow, R., et al., *Inhibitory effects of laser irradiation on peripheral Mammalian nerves and relevance to analgesic effects: a systematic review*. Photomedicine and laser surgery, 2011. **29**(6): p. 365-81.
37. Chow, R.T., M.A. David, and P.J. Armati, *830 nm laser irradiation induces varicosity formation, reduces mitochondrial membrane potential and blocks fast axonal flow in small and medium diameter rat dorsal root ganglion neurons: implications for the analgesic effects of 830 nm laser*. J Peripher Nerv Syst, 2007. **12**(1): p. 28-39.
38. Yan, W., R. Chow, and P.J. Armati, *Inhibitory effects of visible 650-nm and infrared 808-nm laser irradiation on somatosensory and compound muscle action potentials in rat sciatic nerve: implications for laser-induced analgesia*. Journal of the peripheral nervous system : JPNS, 2011. **16**(2): p. 130-5.
39. Artes-Ribas, M., J. Arnabat-Dominguez, and A. Puigdollers, *Analgesic effect of a low-level laser therapy (830 nm) in early orthodontic session*. Lasers in medical science, 2012.
40. Carrasco, T.G., et al., *Evaluation of low intensity laser therapy in myofascial pain syndrome*. Cranio : the journal of craniomandibular practice, 2009. **27**(4): p. 243-7.
41. Bjordal, J.M., et al., *A systematic review of low level laser therapy with location-specific doses for pain from chronic joint disorders*. Aust J Physiother, 2003. **49**(2): p. 107-16.
42. Chen, K.H., et al., *Electrophysiologic effects of a therapeutic laser on myofascial trigger spots of rabbit skeletal muscles*. American journal of physical medicine & rehabilitation / Association of Academic Physiatrists, 2008. **87**(12): p. 1006-14.
43. Snyder-Mackler, L., et al., *Effects of helium-neon laser irradiation on skin resistance and pain in patients with trigger points in the neck or back*. Physical therapy, 1989. **69**(5): p. 336-41.

44. Huang, Y.Y., et al., *Biphasic dose response in low level light therapy*. Dose Response, 2009. **7**(4): p. 358-83.
45. Huang, Y.Y., et al., *Biphasic dose response in low level light therapy - an update*. Dose-response : a publication of International Hormesis Society, 2011. **9**(4): p. 602-18.
46. Sommer, A.P., et al., *Biostimulatory windows in low-intensity laser activation: lasers, scanners, and NASA's light-emitting diode array system*. J Clin Laser Med Surg, 2001. **19**(1): p. 29-33.
47. Jenkins, P.A. and J.D. Carroll, *How to report low-level laser therapy (LLLT)/photomedicine dose and beam parameters in clinical and laboratory studies*. Photomedicine and Laser Surgery, 2011. **29**(12): p. 785-7.
48. Tumilty, S., et al., *Low level laser session of tendinopathy: a systematic review with meta-analysis*. Photomed Laser Surg, 2010. **28**(1): p. 3-16.
49. Bjordal, J.M., et al., *A systematic review with procedural assessments and meta-analysis of Low Level Laser Therapy in lateral elbow tendinopathy (tennis elbow)*. BMC Musculoskeletal Disorders, 2008. **9**: p. 75.
50. Smith, K., *The photobiological basis of low level laser radiation therapy*. Laser Therapy, 1991. **3**: p. 19-24.
51. Byrnes, K.R., et al., *Light promotes regeneration and functional recovery and alters the immune response after spinal cord injury*. Lasers in surgery and medicine, 2005. **36**(3): p. 171-85.
52. Igic, M., et al., *Cytomorphometric and clinical investigation of the gingiva before and after low-level laser therapy of gingivitis in children*. Lasers in medical science, 2012. **27**(4): p. 843-8.
53. Martu, S., et al., *Healing process and laser therapy in the superficial periodontium: a histological study*. Romanian journal of morphology and embryology = Revue roumaine de morphologie et embryologie, 2012. **53**(1): p. 111-6.
54. Kim, S.J., et al., *Effects of low-intensity laser therapy on periodontal tissue remodeling during relapse and retention of orthodontically moved teeth*. Lasers in medical science, 2012.
55. Aimbire, F., et al., *Low-level laser therapy induces dose-dependent reduction of TNFalpha levels in acute inflammation*. Photomed Laser Surg, 2006. **24**(1): p. 33-7.
56. Pejicic, A., et al., *The effects of low level laser irradiation on gingival inflammation*. Photomedicine and Laser Surgery, 2010. **28**(1): p. 69-74.
57. Lievens, P., *The influence of laser irradiation on the motricity of lymphatical system and on the wound healing process*. Proceedings of the International Congress on Laser in Medicine and Surgery, Bologna, Italy, 1986: p. 4.
58. Esper, M.A., R.A. Nicolau, and E.A. Arisawa, *The effect of two phototherapy protocols on pain control in orthodontic procedure--a preliminary clinical study*. Lasers in medical science, 2011. **26**(5): p. 657-63.
59. Bjordal, J.M., et al., *A systematic review with meta-analysis of the effect of low-level laser therapy (LLLT) in cancer therapy-induced oral mucositis*. Supportive care in cancer : official journal of the Multinational Association of Supportive Care in Cancer, 2011.
60. Markovic, A.B. and L. Todorovic, *Postoperative analgesia after lower third molar surgery: contribution of the use of long-acting local anesthetics, low-power laser, and diclofenac*. Oral surgery, oral medicine, oral pathology, oral radiology, and endodontics, 2006. **102**(5): p. e4-8.
61. Aras, M.H. and M. Gungormus, *The effect of low-level laser therapy on trismus and facial swelling following surgical extraction of a lower third molar*. Photomedicine and Laser Surgery, 2009. **27**(1): p. 21-4.
62. Moore, K.C., et al., *The effect of infrared laser irradiation on the duration and severity of postoperative pain a double blind trial*. Laser Therapy, 1992. **Vol 4**: p. 5.
63. Ando, T., et al., *Comparison of therapeutic effects between pulsed and continuous wave 810-nm wavelength laser irradiation for traumatic brain injury in mice*. PLoS ONE, 2011. **6**(10): p. e26212.
64. Hashmi, J.T., et al., *Effect of pulsing in low-level light therapy*. Lasers in surgery and medicine, 2010. **42**(6): p. 450-66.
65. Oron, U., et al., *Attenuation of infarct size in rats and dogs after myocardial infarction by low-energy laser irradiation*. Lasers in surgery and medicine, 2001. **28**(3): p. 204-11.

66. Lanzafame, R.J., et al., *Reciprocity of exposure time and irradiance on energy density during photoradiation on wound healing in a murine pressure ulcer model*. *Lasers in surgery and medicine*, 2007. **39**(6): p. 534-42.
67. Castano, A.P., et al., *Low-level laser therapy for zymosan-induced arthritis in rats: Importance of illumination time*. *Lasers in surgery and medicine*, 2007. **39**(6): p. 543-50.
68. Fred M. Dickey, S.C.H., *Laser Beam Shaping: Theory and Techniques*. 2000.
69. Salmos-Brito, J.A., et al., *Evaluation of low-level laser therapy in patients with acute and chronic temporomandibular disorders*. *Lasers in medical science*, 2012.
70. Marini, I., M.R. Gatto, and G.A. Bonetti, *Effects of superpulsed low-level laser therapy on temporomandibular joint pain*. *The Clinical journal of pain*, 2010. **26**(7): p. 611-6.
71. Mazzetto, M.O., T.H. Hotta, and R.C. Pizzo, *Measurements of jaw movements and TMJ pain intensity in patients treated with GaAlAs laser*. *Brazilian dental journal*, 2010. **21**(4): p. 356-60.